A NEW ANTIVIRAL SUBSTANCE S-15-1, STREPTOTHRICIN GROUP ANTIBIOTIC

KEI ARIMA, TAKESHI KAWAMURA and TERUHIKO BEPPU

Laboratory of Microbiology, Department of Agricultural Chemistry, University of Tokyo, Bunkyo-ku, Tokyo, Japan

(Received for publication January 7, 1972)

In the screening for antiviral antibiotics produced by *Streptomyces* soil isolates, several strains were found to produce cytotoxic and/or antiviral antibiotics. One antiviral antibiotic designated S-15-1 was isolated from soil isolate *Streptomyces* S-15-1. The antibiotic S-15-1 shows noncytotoxic *in vitro* inhibitory activity against Newcastle disease virus, and it has also inhibitory activity against gram-positive, gram-negative bacteria as well as some yeasts.

In the course of an antiviral antibiotic screening program utilizing a paper-disc agar diffusion technique¹⁾ involving Newcastle disease virus (NDV) MIYADERA strain²⁾ with a primary culture of chick embryo fibroblast (CEF), it was found that 62 strains of unidentified *Streptomyces* obtained from soil samples produced cytotoxic and/or antiviral antibiotics. One of the isolates, *Streptomyces* S-15-1, produced a novel watersoluble basic antibiotic. This noncytotoxic antibiotic, designated S-15-1, was active *in vitro* against NDV and active against gram-positive, gram-negative bacteria, and some strains of yeast.

This paper describes the screening for antiviral antibiotics produced by one isolate of *Streptomyces*, and also the production, isolation, purification, and physico-chemical and biological properties of S-15-1.

Screening of the Antiviral Antibiotics

Approximately one thousand strains of *Streptomyces* obtained from soil samples were inoculated in each of two media and cultured at 26.5°C for 5 days with shaking. The media were as follows: medium A contained 3.0 % glycerol, 1.0 % peptone, 0.5 % $(NH_4)_2HPO_4$, 1.0 % K_2HPO_4 , 0.5 % NaCl, 0.05 % $MgSO_4 \cdot 7H_2O$, and 0.001 % $FeSO_4 \cdot 7H_2O$ (pH 7.0); medium B contained 3.0 % starch, 1.0 % peptone, 0.4 % $NaNO_3$, 0.2 % K_2HPO_4 , 0.1 % $MgSO_4 \cdot 7H_2O$, and 0.002 % $FeSO_4 \cdot 7H_2O$ (pH 7.0).

After cultivation an equal volume of acetone was added to the whole broth, and antiviral activity in supernatant of broth was examined by the method of HERRMANN and his coworkers, and at the same time antimicrobial activity was also determined. In all, 47 strains produced only the cytotoxic and/or antiviral antibiotics and 15 strains produced the cytotoxic and/or antiviral as well as antimicrobial activity. On the other hand, 126 strains of *Streptomyces* produced the antimicrobial antibiotics.

Table 1 indicates the strains which produced the cytotoxic and/or antiviral antibiotics. Cytotoxic activity was observed more frequently than antiviral activity.

Org	anisms	Me- dium	CTZ	PIZ	Org	anisms	Me-	CTZ	PIZ	Org	anisms	Me-	CTZ	PIZ	Org	anisms	Me- dium	стΖ	PIZ
S	1-2	B	19		S	6-3	B	22		S	12-6	B	12		S	16-22	B	13	
ŝ	2-2	B	18		ŝ	6-4	B	12		s	12-11	B	12	_	s	16-32	B	_	16
S	2-3	В	18	_	S	6-5	A	11		S	12-18	В	20	_	S	16-33	В	9	11
S	2-4	В	-	12	S	7-1	В		8	S	12-21	В	12		S	17-1	В	17	
S	2-8	В	10		S	8-10	В	26	_	S	13-2	В	8		S	17-3	В	_	11
S	2-10	В		18	S	8-11	В	25	******	S	13 - 22	В	_	12	S	17-5	В	10	—
S	2-12	В	15	—	S	8-12	В	10	•	S	13-23	В	-	10	S	17-7	В	8	_
S	2-14	В	20		S	9–9	В	24		S	14-4	В	16	18	S	18-2	В	22	
S	2-16	Α		7	S	9-10	в	23	_	S	16-11	в	—	10	S	19-4	В	7	
S	2-18	в	11	17	S	10-4	В	18		S	16 - 12	В	20		S	19-6	В	18	
S	4-4	в	19	—	S	10-6	Α	—	10	S	16 - 13	В	14	_	S	19-31	В	—	15
S	5-4	В	-	14	S	10-11	В	10	14	S	16-21	В	10						

Table 1. Antiviral and cytotoxic activity of the antibiotics produced by *Streptomyces* obtained from soil samples

Monolayers of chick embryo fibroblast formed on Petri dish (90 mm in diameter) using the E.C. HERRMAN, Jr., *et al.* method were infected by NDV MIYADERA strain and overlayed with soft agar medium. Paper discs impregnated with samples (Toyo Roshi Co., 6 mm in diameter) were placed on the agar overlay and plates were incubated at 37°C. After 48-hour incubation, diameters of plaque-free protected zones (PIZ) and cytotoxic zones (CTZ) were measured and expressed in mm.

Table	2.	Antivi	ral	and	cyt	otoxic	activity	and	antimicrol	bia]
		activity	of	the	anti	ibiotics	produce	d by	Streptomy	ces
		obtained	fr	om s	soil	sample	es			

Orga-		Midio	CT7	DI7	Growth inhibition zone								
n	isms	wituta	CIL	F IZ	S. lutea	B. subtilis	E. coli	C. albicans					
S	2-1	В	13		14.5	_		12.0					
S	2-5	В	12	_	12.0			10.0					
S	2-6	В	11	16	-	_	_	15.0					
S	5 - 2	Α	12	16	19.8		_	_					
S	6 - 1	В		9	16.6	13.6		19.8					
S	6-8	В	19		20.2	27.5	_	10.7					
S	8-14	В	16	—	_	13.0	—	_					
S	9-2	Α		12	23.5	11.8	—						
S	15-1	В		19	17.4	20.2	10.6	15.0					
S	16 - 1	А, В	17		_	14.3		16.4					
S	17-7	В	24	_	15.0	14.0	—	_					
S	17 - 20	В	30	_	12.0	8.5							
S	17 - 21	В	14		12.5	13.0		—					
S	19–17	В	16			10.0	10.0	_					
S	19-25	В		18		—	12.0	<u> </u>					

A few strains produced high concentrations of cytotoxic activity and small amounts of antiviral activity.

Table 2 shows the strains which produced cytotoxic and/or antiviral and antimicrobial antibiotics. *Streptomyces* isolated from soil samples grow very poor on medium A, and some strains produce lower concentrations of antibiotics in this medium (Tables 1 and 2).

The NDV MIYADERA strain and CEF monolayer system was used. Diameters of plaque-free protected zone (PIZ) and cytotoxic zone (CTZ) were measured and expressed in mm. The paper disc used was 6 mm in diameter for broth antiviral and cytotoxic activity, and 8 mm in diameter for antimicrobial activity (Toyo Roshi Co.).

Production, Isolation, and Purification of S-15-1

Strain S-15-1 was of particular interest because of produced noncytotoxic antiviral activity with very broad spectrum antimirobial activity. Streptomyces S-15-1 was grown in 20 liters of medium, consisting of 3.0% starch, 1.0% peptone, 0.4% NaNO₃, 0.2% K₂HPO₄, 0.1% KCl, 0.1% MgSO₄·7H₂O, and 0.002% FeSO₄·7H₂O (pH 7.0), sterilized at 120°C for 20 minutes in a 30-liter stainless steel jar fermentor. Each jar was inoculated with 5.0% (v/v) of 48-hour-old seed culture grown in the same medium in shaker flasks at 26.5°C. The fermentation was carried out in a jar fermentor for 55 hours at 26.5°C, agitating 280 rpm, with 100% aeration. The

final pH of the culture broth was 7.8. Fermented medium (40 liters) was adjusted to pH 6.0 with diluted hydrochloric acid, and filtered to remove cell debris. The culture filtrate was adjusted to pH 7.2 and stirred with active carbon (5%, v/v). After the filtration of carbon, the carbon cake was washed twice with water and the active components were eluted with 50% aqueous methanol containing 0.1% hydrochloric acid. The active eluates were neutralized by passage through a Dowex-44 (OH⁻ type) column and they were concentrated under reduced pressure at 30°C. The concentrated solution was lyophilized, the crude powder was dissolved in methanol, the insoluble residue was separated by filtration, and the clear methanol solution was concentrated under reduced pressure. The S-15-1 antibiotic was precipitated as the hydrochloride by addition of acetone. The precipitate was separated, washed with acetone and dried.

Futher purification was done by a cellulose column with *n*-propanol-pyridineacetic acid-water (15:10:3:12) solvent system. Active fractions were concentrated, and the antibiotic acetate was converted to the sulfate salt utilizing an Amberlite IRC-50 (H⁺ type) column. The solution was concentrated under reduced pressure at 30°C and lyophilized. Two hundred milligrams of S-15-1 sulfate were thus obtained as a white powder. The purified S-15-1 sulfate was detected in thin-layer chromatograms as a clear single spot by ninhydrin tests and by antiviral and antimicrobial tests. Cellulose thin-layer chromatograms were developed with *n*-propanol-pyridineacetic acid-water (15:10:3:12) and *n*-butanol-acetic acid-water (2:1:1). Silica gel plates were developed with chloroform-methanol-17% aqueous ammonia (2:1:1, upper layer) and pyridine-acetic acid-water (50:35:15).

Physico-chemical and Biological Properties of S-15-1

S-15-1 sulfate is soluble in water, insoluble in methanol, ethanol, acetone, ethylacetate, chloroform, ethylether, benzene, and hexane. It is stable at powder state and at 90° C for 10 minutes with the solution of pH 2.0, but little activity remains when it is treated at pH 8.0 and 90° C for 10 minutes.

It melts at $161 \sim 171^{\circ}$ C under decomposition. It shows only end absorption in the ultraviolet spectrum. The infrared absorption spectrum in KBr disc is presented in Fig.1. It has characteristic absorption bands at the following frequencies: 3400, 1718, 1655, 1615, 1559, 1495, 1395, 1335, 1315, 1250, 1220, 1193, 1080, 1055, 1005, 890, and 780 cm⁻¹.



Fig. 1. Infrared spectrum of S15-1 sulfate in KBr-disc.

Table	3.	Thin-layer	chromatography	of	S-15-1	sulfate
A)	Cel	lulose plate				

•					
Solvent systems	Rf values				
n-Propanol – pyridine – acetic acid – water (15:10:3:12)					
n-Butanol - acetic acid - water $(2:1:1)$	0.15				
Wet <i>n</i> -but anol containing 2 % (w/v) <i>p</i> -toluene-sulfonic acid					
B) Silica gel G plate					
Solvent systems	Rf values				
Chloroform - methanol - 17 % aqueous ammonia (2:1:1) (upper layer)	0. 28				
n-Butanol – acetic acid – water $(2:1:1)$	0.05				

Table 4. Antiviral activity of S15-1 sulfate by the agar diffusion method

Concentration (mcg/ml)	PIZ
2,000	27
100	19
80	15
30	10
10	-

The optical rotation of the S-15-1 hydrochloride is $[\alpha]_D - 26^\circ$ (c 1.0, water). Elemental analysis is as follows:

Calcd. for $C_{22}H_{49}N_9O_{11}\cdot 2HCl$:

C 38.37, H 7.41, N 18.31, O 25.58, Cl 10.32. Found:

C 38.30, H 7.30, N 18.80, O 24.50, Cl 10.40.

The antibiotic is positive to MOLISCH, SAKAGUCHI (strong), ELSON-MORGAN (pink), FEHLING, anthrone, maltol (weak) and ninhydrin tests, but negative to biuret and orcinol tests. The results of thin-layer chromatography are shown in Table 3.

The antiviral activity of the antibiotic is shown in Table 4. No cytotoxicity was observed at the higher concentration. The antimicrobial activities were examined using the agar streak dilution method and the results are shown in Table 5. It shows inhibitory activity against gram-positive and gram-negative bacteria, and some yeast strains. The intravenous LD_{50} in mice (ddY) Diameter of plaque-free inhibition zones (PIZ) were measured and expressed in mm.

Table 5. Antimicrobial activity of S15-1 sulfate

Test organisms*	M.I.C. (mcg/ml)
Staphylococcus aureus FDA 209P	0.78
Staphylococcus aureus	>50.00
FDA 209P (ST-R, SM-R)	
Sarcina lutea PCI 1001	6.25
Bacillus subtilis PCI 219	0.78
Bacillus cereus IFO 3001	25.00
Proteus vulgaris IFO 3851	1.56
Escherichia coli K-12	3.13
Escherichia coli B (SM-R)	3.13
Escherichia coli (KM-R)	3.13
Escherichia coli W 3630	1.56
(SA, PC, TC, CP, SM-R)	
Escherichia coli (ST-R)	>50.00
Pseudomonas aeruginosa IFO 3448	>50.00
Mycobacterium 607 (SM-R)	6.25
Saccharomyces cerevisiae	>50.00
Hansen Kyokai 6	
Candida albicans IAM 4888	50.00
Cryptococcus neoformans	25.00
IAM 4514	
Penicillium chrysogenum Тном FAT-917	>50. 00
Aspergillus niger van Thieghen	50.00

* -R: resistant.

ST: streptothricin, SM: streptomycin, KM: kanamycin, SA: sulfonamide, PC: penicillin G, TC: tetracycline, CP: chloramphenicol.

is 150 mg/kg. However, as occurs with streptothricin, delayed toxicity begins to appear after 2 days at 100 mg/kg, and after 3 days at 50 mg/kg.

Discussion

All *Streptomyces* isolated were tested for cytotoxic, antiviral, and antimicrobial activity. Many strains produced cytotoxic antibiotics, but few strains produced only antiviral antibiotics. One isolate, *Streptomyces* S-15-1, produced a noncytotoxic, antiviral antibiotic with broad antimicrobial activity. This water-soluble basic antibiotic was differentiated from the known antibiotics by its physico-chemical and biological properties. The antibiotic gave positive SAKAGUCHI (strong), maltol (weak), nynhydrin, Molisch, and anthrone reactions, and negative orcinol and biuret reactions. It was differentiated from streptomycin group antibiotics, because of a negative ninhydrin reaction and its antiviral activity and the activity against *Candida albicans*.

The positive SAKAGUCHI and maltol reactions, and negative orcinol reaction differentiate the antibiotic from kanamycins, neomycins, paromomycins, zygomycins, hydroxymycins, and catenulin.

It was also differentiated from streptothricin group antibiotics (Tables 6 and 7). New streptothricin-like antibiotics which were recently found and reported by various groups have higher Rf values than that of kanamycin on silica gel thin-layer chromatogram developed with the upper layer from a mixture of chloroform – methanol – 17 % aqueous ammonia (2:1:1). Antibiotic S-15-1 has a much smaller Rf value than that of kanamycin. Strepto- Table 7.

that of kanamycin. Streptothricins and racemomycins gave negative SAKAGUCHI and maltol reactions.

Antiviral antibiotics such as amidinomycin, phagolessin, phagocidin, grasseriomycin, and myxoviromycin were differentiated by color reaction and antimicrobial activity. These antiviral antibiotics are maltol negative. Grasseriomycin is inactive against Candida albicans. Amidinomycin, phagolessin, phagocidin, myxoviromycin, and amidinomycin are weakly active

					1	XI VA	anues			
A 4 ¹ b ¹ c 4 ¹ c -			A				I	3		С
Antibiotics		3	rlC			PPC	TLC	PPC		TLC
	Ι		Π	II	I		I			III
S 15-1	0.05	0.	08	0.5	0	0.10	0.15	0.00		0.28
Boseimycin ³⁾				0.7	5			0. 08		
BD-12 ⁴⁾						0.54		0. 29		0.78
BY-814)		0.	36^{12}			0. 45		0.17		0.52^{12}
Citromycin ¹²⁾		0.	35							0. 52
LL-AC54112)		0.	35			0. 50				0.52
Racemomycin		0.	25			0. 38	0.20			0.36
$Sclerothricin^{8)}$										₀ . 78
SF-70111)	0.66	0.	3312)				0.23			0.72
Streptothricin	0.26			0.7	2	0. 38				0.26
Viomycin	0.21					0.28				$0.\ 11^{13}$
Yazumycin ¹¹⁾		ļ		0.6	4			0. 09	0.	$3 \sim 0.33$
A: n-Propano	l-py	rio	line -	ace	ti	c aci	d - wa	ter ((15:	10:3:

B: Wet n-butanol containing 2.0% (w/v) p-toluenesulfonic acid

C: Chloroform-methanol-17 % aqueous ammonia (2:1: 1) (upper layer)

I: Cellulose MN 300 powder

II: Avicel SF plate III: Silica gel G

e 7. Comparison of S15-1 with other known antibiotics

A 4 ! Is ! = 4 ! = =			Colo	r reactio	actions ISCH Ninhydrin + + - + - + (in pyridine) - + + - + + - + + - + + - + + - + + + - + + + + + + + + + + + + + +	
Antibiotics	Biuret	FEHLING	Maltol	Molisch	Ninhydrin	Saka- guchi
S 15-1	_	+	+	+	+	+
Boseimycin ³⁾	+	-		+	+	+
BD-124)	?	?	-	—	+	
BY-81 ⁴⁾		?	_	_	+	-
Citromycin ⁵⁾		_	_		+	
					(in pyridine)	
EC-749 C ⁶⁾		-			—	—
LL-AC 5417)	+	+		_		—
Racemomycin		+	-		+	
Sclerothricin ⁸⁾	—		-	_	+	±
SF-7019)		+			+	
Streptothricin	+	+		—	+	
Viomycin ¹⁰⁾	+	+-	_		+	+
Yazumycin ¹¹⁾	+	+			+	+
	(weak)					

+: Positive -: Negative ±: Slightly colored ?: Doubtful

against gram-positive and gram-negative bacteria.

Accordingly it can be concluded that the antibiotic S-15-1 is a new antiviral and antimicrobial antibiotic.

Acknowledgement

The authors express their sincere thanks to Dr. Z. ABE, Dr. A. NAGATA, Dr. K. HAYANO, Dr. T. ANDO, and Dr. K. MIZUNO, Research Laboratory, Toyo Jozo Co., Ltd., for their kind supply of resistant strains and for the toxicity tests.

Table 6. Chromatographic comparison of S15-1 with other known antibiotics

nc

References

- HERRMANN, E. C., Jr.; J. GABLIKS, C. ENGLE & P. L. PERLMAN: Agar diffusion method for detection and bioassay of antiviral antibiotics. Proc. Soc. Expt. Biol. Med. 103: 625~628, 1960
- ARIMA, K.; A. TAKATSUKI, S. SUZUKI, K. ANDO & G. TAMURA : Antiviral activity of trichothecin. J. Antibiotics 21 : 158~159, 1968
- SINHA, R. K. : Boseimycin a new streptothricin-like antibiotic. J. Antibiotics 23: 360~364, 1970
- ITO, Y.; Y. OHASHI, Y. SAKURAI, M. SAKURAZAWA, H. YOSHIDA, S. AWATAGUCHI & T. OKUDA: New basic water-soluble antibiotics BD-12 and BY-81. II. Isolation, purification and properties. J. Antibiotics 21: 307~312, 1968
- KUSAKABE, Y.; Y. YAMAUCHI, C. NAGATSU, H. ABE, K. AKASAKI & S. SHIRATO: Citromycin, a new antibiotic. I. Isolation and characterization. J. Antibiotics 22: 112~118, 1969
- SHOJI, J.; S. KOZUKI, M. EBATA & H. OTSUKA: A water-soluble basic antibiotic E-749C identical with LL-AC541. J. Antibiotics 21: 509~511, 1968
- 7) ZEINOVSKY, V.; W. K. HAUSMANN, E. R. WETZEL, D. B. BORDERS & E. L. PATTERSON: Isolation and characterization of antibiotic LL-AC 541. Appl. Microbiol. 16:614~616, 1968
- KONO, Y.; S. MAKINO, S. TAKEUCHI & H. YONEHARA: Sclerothricin, a new basic antibiotic. J. Antibiotics 22: 583~589, 1969
- 9) TSURUOKA, T.; T. SHOUMURA, N. EZAKI, T. NIWA & T. NIIDA: SF-701, a new streptothricinlike antibiotic. J. Antibiotics 21: 237~238, 1968
- HASKELL, T. H.; S. A. FUSARI, R. P. FROHARDT & Q. R. BARTZ: The chemistry of viomycin. J. Am. Chem. Soc. 74: 599~602, 1952
- AKASAKI, K.; H. ABE, A. SEINO & S. SHIRATO: Yazumycin, a new antibiotic produced by Streptomyces lavendulae. J. Antibiotics 21: 98~105, 1968
- TANIYAMA, H. & Y. SAWADA : The identity of citromycin with LL-AC541, E-749-C, and BY-81. J. Antibiotics 24: 708~710, 1971
- 13) IKEKAWA, T.; F. IWAMI, E. AKITA & H. UMEZAWA: Application of thin-layer chromatography for separation and identification of antibiotics. J. Antibiotics, Ser. A 16: 56~57, 1963
- 14) KORZYBSKI, T.; Z. KOWSZYK-GRINDIFER & W. KURYŁOWICZ: Antibiotics; Vol. 1, Origin, nature and properties. Pergamon Press (Oxford, London, Edinburgh, New York, Tronto, Sydney, Paris and Braunschweig). PWN-Polish Scientific Publisher (Warszawa), 1967